Serial No.: 10/057,467

Filed: January 22, 2002

changes made to the specification and claims by the current amendment. The attached page is captioned "Version with markings to show changes made."

Entry of this amendment is respectfully requested. The amendments are made in adherence with 37 C.F.R. § 1.821-1.825. This amendment is accompanied by a Request to Use Computer Readable Form of Sequence Listing From Another Application and a paper copy of the sequence information from that prior application. This amendment contains no new matter. Applicant submits that this amendment, the accompanying computer readable sequence listing, and the paper copy thereof serve to place this application in a condition of adherence to the rules 37 C.F.R. § 1.821-1.825.

Please direct any calls in connection with this application to the undersigned at (415) 781-1989.

Respectfully submitted,

DORSEY & WHITNEY LLP

Dated:

May 6, 2002

Four Embarcadero Center

Suite 3400

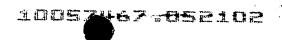
San Francisco, CA 94111-4187

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Robin M. Silva, Reg. No. 38,304

Filed under 37 C.F.R. Section 1.34(a)

-6-



VERSION WITH MARKINGS TO SHOW CHANGES MADE

IN THE SPECIFICATION:

Paragraph beginning at page 3, line 22, has been amended as follows:

— Figure 2. Creation of a library of random peptides in a retrovirus DNA construct by primed DNA synthesis (SEQ ID NOS:10-14).—

Paragraph beginning at page 4, line 9, has been amended as follows:

- The introduced nucleic acids and resultant expression products are randomized, meaning that each nucleic acid and peptide consists of essentially random nucleotides and amino acids, respectively. The library may be fully random or biased, e.g., in nucleotide/residue frequency generally or per position. For example, a biased library may encode peptides for interactions with known classes of molecules, such as SH-3 domain proteins, as defined by peptides containing XXXPPXPXX (where X=randomized residues; SEQ ID NO:1). In other embodiments, the nucleotides or residues are randomized within a defined class, e.g., of hydrophobic amino acids, of purines, etc. In any event, where the ultimate expression product is a nucleic acid, at least 10, preferably at east 12, more preferably at least 15, most preferably at least 21 nucleotide positions need to be randomized; more if the randomization is less than perfect. Similarly, at least 5, preferably at least 6, more preferably at least 7 amino acid positions need to be randomized; again, more if the randomization is less than perfect.—

Paragraph beginning at page 9, line 26, has been amended as follows:

-7-

1081286

- A scheme for generating a peptide library in the pBabe Puro vector is shown in Figure 2. Primers for PCR were synthesized, purified and deprotected according to standard protocols. Primer 1, complementary to polylinker sequences in the pBabe Puro retroviral construct, has the sequence 5' GCT TAG CAA GAT CTC TAC GGT GGA CCK NNK NNK NNK NNK NNK NNK NNK NNK NNC CCC ACT CCC ATG GTC CTA CGT ACC ACC ACA CTG GG 3' (SEQ ID NO:2). N represents any of the four bases; K is limited to G or T. Primer 2 has the sequence 5' GCT TAG CAA GAT CTG TGT GTC AGT TAG GGT GTG G 3' (SEO ID NO:3) and is complementary to sequences within the pUC18 origin of replication. PCR was carried out for 8 rounds using primer 1, primer 2, Babe Puro as template, and a mixture of Taq DNA Polymerase (Promega) and Deep Vent DNA Polymerase (New England Biolabs) in a ratio of 128 Taq: 1 Deep Vent as described in Barnes (1994) Proc. Natl. Acad. Sci. USA, 91, pp. 2216-2220. The amplified PCR product was purified, digested with restriction enzymes Bgl II and Not I (Promega), purified again and ligated with the corresponding Bam HI-Not I fragment of pBabe Puro. After transformation the resulting library contained $\sim 2x10^8$ clones, greater than 80% of which contained inserts. -

Paragraph beginning at page 10, line 12, has been amended as follows:

— Oligonucleotides were synthesized and purified according to standard protocols. The "library" oligonucleotides have the sequence 5' CTG GAG AAC CAG GAC CAT GGG C (NNK)₁₀ GGG CCC CCT TAA ACC ATT AAA T 3' (SEQ ID NO:4) or 5' CTG GAG AAC CAG GAC CAT GGG CNN KNN KNN KCC TCC

anetha a ann a amhraigh an seige go go go go go ann a dhaanna a go go garainn a ann a an ann an ann an ann an a

CNN KCC TNN KNN KGG GCC CCC TTA AAC CAT TAA AT 3'(SEQ ID NO:5). A third oligonucleotide ("constant"), complementary to the 3' ends of the library oligonucleotides, has the sequence 5'TCA TGC ATC CAA TTT AAT GGT TTA AG 3'(SEQ ID NO:6). As shown in Fig. 3 Fig. 2, each library oligonucleotide is annealed to the constant oligonucleotide, converted to double stranded DNA with Sequenase (United States Biochemical) or Klenow (Promega), digested with restriction enzyme Bst XI (New England Biolabs), and purified and ligated with the appropriate Bst XI-digested retroviral construct. Transformation efficiencies are ~ 2 x 10⁸ clones per microgram of ligated DNA, greater than 90% of which contain an insert. A representative retrovirus is shown in Fig. 4; see also, retroviral mucleotide sequence below: vector with presentation construct nucleotide sequence (SEQ ID NO:7). —

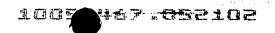
Paragraph beginning at page 10, line 24, has been deleted as follows:

- Retroviral vector with presentation construct:

TGAAAGACCCCACCTGTAGGTTTGGCAAGCTAGCTTAAGTAACGCCATTTT
GCAAGGCATGGAAAATACATAACTGAGAATAGAGAAGTTCAGATCAAGG
TTAGGAACAGAGAGACAGCAGAATATGGGCCAAACAGGATATCTGTGGT
AAGCAGTTCCTGCCCCGGCTCAGGGCCAAGAACAGATGTTCCCAGGGTGC
GGTCCCGCCCTCAGCAGTTTCTAGAGAACCATCAGATGTTTCCAGGGTGC
C

CCAAGGACCTGAAAATGACCCTGTGCCTTATTTGAACTAACCAATCAGTT
CGCTTCTCGCTTCTGCTCCCCGAGCTCAATAAAAGAG
CCCACAACCCCTCACTCGGCGCGCCCAGTCCTCCGATAGACTGCGTCGCCC

1081286 -9-



GGCTACCCGTATTCCCAATAAAGCCTCTTGCTGTTTGCATCCGAATCGTGG ACTCCCTGATCCTTGGGAGGGTCTCCTCAGATTGATTGACTGCCCACCTCG GGGTCTTTCATTTGGAGGTTCCACCGAGATTTGGAGACCCCTGCCTAGGG ACCACCGACCCCCCCCCGGGAGGTAAGCTGGCCAGCGGTCGTTTCCTGT CTGTCTCTCTTTGTGCGTGTTTGTGCCGGCATCTAATGTTTGCGCCTGCG TCTGTACTAGTTAGCTAACTAGCTCTGTATCTGGCGGACCCGTGGTGGAAC TGACGAGTTCTGAACACCCGGCCGCAACCCTGGGAGACGTCCCAGGGACT TTGGGGCCGTTTTTGTGGCCCGACCTGAGGAAGGGAGTCGATGTGGAAT **CCGACCCGTCAGGATATGTGGTTCTGGTAGGAGACGAGAACCTAAAACA** GTTCCCCCTCCGTCTGAATTTTTGCTTTCGGTTTGGAACCGAAGCCGCGC GTGTTTCTGTATTTGTCTGAAAATTAGGGCCAGACTGTTACCACTCCCTTA AGTTTGACCTTAGGTCACTGGAAAGATGTCGAGCGGATCGCTCACAACCA GTCGGTAGATGTCAAGAAGAGACGTTGGGTTACCTTCTGCTCCAGAAT GGCCAACCTTTAACGTCGGATGGCCGCGAGACGCCACCTTTAACCGAGAC CTCATCACCCAGGTTAAGATCAAGGTCTTTTCACCTGGCCCGCATGGACAC CCAGACCAGGTCCCCTACATCGTGACCTGGGAAGCCTTTGGCTTTTGACCCC **CCTCCCTGGGTCAAGCCCTTTGTACACCCTAAGCCTCCGCCTCCTCTTCCT** CCATCCCCCGTCTCTCCCCCTTGAACCTCCTCGTTCGACCCCGCCTCGA TCCTCCTTTATCCAGCCCTCACTCCTTCTCTAGGCGCCGGAATTCCAGGA CCATGGCGGCCCCCTTAAACCATTAAATTGGTAAAATAAAGGATCCGT

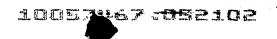
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AAGTAACGCCATTTTGCAAGGCATGGAAAATACATAACTGAGAATAGAGA AGTTCAGATCAAGGTTAGGAACAGAGAGACAGCAGAATATGGGCCAAAC AGGATATETGTGGTAAGEAGTTEETGEEECGGCTCAGGGCCAAGAACAGA TGGTCCCAGATGCGGTCCCGCCCTCAGCAGTTTCTAGAGAACCATCAGA TGTTTCCAGGGTGCCCCAAGGACCTGAAAATGACCCTGTGCCTTATTTGAA CTAACCAATCAGTTCGCTTCTCGCTTCTGCTCCCCGA GCTCAATAAAAGAGCCCACAACCCCTCACTCGCCGCCAGTCCTCCGAT AGACTGCGTCGCCGGGTACCCGTGTATCCAATAAACCCTCTTGCAGTTGC ATCCGACTTGTGGTCTCGCTGTTCCTTGGGAGGGTCTCCTCTGAGTGATTG ACTACCGTCAGCGGGGTCTTTCATTCGTAATCATGGTCATAGCTGTTTC CTGTGTGAAATTGTTATCCGCTCACAATTCCACACACATACGAGCCGGA AGCATAAAGTGTAAAGCCTGGGGTGCCTAATGAGTGAGCTAACTCACATT AATTGCGTTGCGCTCACTGCCCGCTTTCCAGTCGGGAAACCTGTCGTGCCA **GCTGCATTAATGAATCGGCCAACGCGGGGGGGGGGGGGTTTGCGTATTG** GGCGCTCTTCCGCTCGCTCACTGACTCGCTCGGTCGTTCGGC TGCGCCAGCGTATCAGCTCACTCAAAGGCCGTAATACGGTTATCCACA GAATCAGGGGATAACGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAA AGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATAGGCTC **CGCCCCTGACGAGCATCACAAAAATCGACGCTCAAGTCAGAGGTGGCG AAACCGACAGGACTATAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCC** TCGTGCGCTCTCCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCT

1081286 -11-

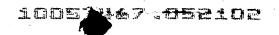
TTCTCCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCTGTAGGTATC TCAGTTCGGTGTAGGTCGTTCGCTCCAAGCTGGGCTGTGTGCACGAACCCC **CCGTTCAGCCGACCGCTGCGCCTTATCCGGTAACTATCGTCTTGAGTCCA** ACCCGTAAGACACGACTTATCGCCACTGGCAGCCACTGGTAACAGG **ATTAGEAGAGEGAGGTATGTAGGEGGTGCTACAGAGTTCTTGAAGTGGTG GCCTAACTACGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTCTGCT** GAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAAC AAACCACCGCTGGTAGCGGTGGTTTTTTTGTTTGCAAGCAGCAGATTACGC **GCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCT** GACGCTCAGTGGAACGAAAACTCACGTTAAGGGATTTTGGTCATGAGATT ATCAAAAGGATCTTCACCTAGATCCTTTTAAATTAAAAAGGATTTTAA ATCAATCTAAAGTATATGAGTAAACTTGGTCTGACAGTTACCAATGCTT AATCAGTGAGGCACCTATCTCAGCGATCTGTCTATTTCGTTCATCCATAGT TGCCTGACTCCCGTCGTGTAGATAACTACGATACGGGAGGGCTTACCAT **CTGGCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCGGCTCCA GATTTATCAGCAATAAACCAGCCAGCCGGAAGGGCCGAGCGCAGAAGTG** GTCCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAG CTAGAGTAAGTAGTTCGCCAGTTAATAGTTTGCGCAACGTTGTTGCCATTG **CCGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCCATGTTGTGCAAA** GCAGTGTTATCACTCATGGTTATGGCAGCACTGCATAATTCTCTTACTGTC

1081286 -12-



ATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCA TTCTGAGAATAGTGTATGCGGCGACCGAGTTGCTCTTGCCCGGCGTCAATA CGGGATAATACCGCGCCACATAGCAGAACTTTAAAAAGTGCTCATCATTGG AAAACGTTCTTCGGGGCGAAAACTCTCAAGGATCTTACCGCTGTTGAGAT CCAGTTCGATGTAACCCACTCGTGCACCCAACTGATCTTCAGCATCTTTTA CTTTCACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCA **AAAAAGGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTCCT** TTTCAATATTGTGAAGCATTTATCAGGGTTATTGTCTCATGAGCGGATA **CATATTTGAATGTATTTAGAAAAATAAACAAATAGGGGTTCCGCGCACAT TTCCCGAAAAGTGCCACCTGACGTCTAAGAAACCATTATTATCATGACA TTAACCTATAAAAATAGGCGTATCACGAGGCCCTTTCGTCTCGCGCGTTTC** GGTGATGACGGTGAAAACCTCTGACACATGCAGCTCCCGGAGACGGTCAC AGCTTGTCTGTAAGCGGATGCCGGGAGCAGACAAGCCCGTCAGGGCGCGT **CAGCGGTGTTGGCGGGTGTCGGGGCTGGCTTAACTATGCGGCATCAGAG CAGATTGTACTGAGAGTGCACCATATGCGGTGTGAAATACCGCACAGATG CCTAAGGAGAAAATACCGCATCAGGCGCCATTCGCCATTCAGGCTGCGCA ACTOTTGGGAAGGGCGATCGGTGCGGGCCTCTTCGCTATTACGCCAGCTG** CCGAAGGGGATGTGCTGCAAGGCGATTAAGTTGGGTAACGCCAGGGTT **TTCCCAGTCACGACGTTGTAAAACGACGCCAGTGCCACGCTCTCCCTTAT GCGACTCCTGCATTAGGAAGCAGCCCAGTAGTAGGTTGAGGCCGTTGAGC** ACCCCCCCCAACGAATGGTGCATGCAAGGAGATGGCGCCCAACAGTC

1081286 -13-



Paragraph beginning at page 13, line 25, has been amended as follows:

— In some embodiments of the invention, expression products are localized to, or preferentially concentrated in, different subcellular compartments within cells, e.g., by using appropriate addition of addressins to a peptide presentation construct, see, Figure 3. Addressins are available for a wide variety of subcellular locales including the nucleus, Golgi, mitochondria, plasma membranes, endoplasmic reticulum, secretory granules, secreted, cell surface (extracellular domain with random), cell surface (intracellular domain random), etc. For example, many proteins whose functions require entry into the cell nucleus include nuclear localization signal (NLS) sequences: generally short, positively charged (basic) domains that serve to direct the entire protein in which they occur to the cell's nucleus. Numerous NLS amino acid sequences have been reported including single basic NLS's such as that of the SV40 (monkey virus) large T Antigen (Pro Lys Lys Lys Arg Lys Val (SEQ ID NO:8), Kalderon (1984), et al., Cell, 39:499-509, and double basic NLS's exemplified by that of the Xenopus (African clawed toad) protein, nucleoplasmin (Ala Val Lys Arg Pro Ala Ala Thr Lys Lys Ala Gly Gln Ala Lys Lys Lys Leu Asp (SEQ ID NO:9), Dingwall, et al., Cell, 30:449-458, 1982 and Dingwall, et al., J. Cell Biol., 107:641-849; 1988). Numerous localization studies have demonstrated that

1081286 -14-

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Serial No.: 10/057,467 **Filed**: January 22, 2002

NLSs incorporated in synthetic peptides or grafted onto reporter proteins not normally targeted to the cell nucleus cause these peptides and reporter proteins to be concentrated in the nucleus. See, for example, Dingwall, and Laskey, Ann, Rev. Cell Biol., 2:367-390, 1986; Bonnerot, et al., Proc. Natl. Acad. Sci. USA, 84:6795-6799, 1987; Galileo, et al., Proc. Natl. Acad. Sci. USA, 87:458-462, 1990.—

On page 16, immediately preceding the claims, the enclosed "SEQUENCE LISTING" was inserted into the specification.

-15-